Comparison of novel magnetic polyaluminum chlorides involved coagulation with traditional magnetic seeding coagulation: Coagulant characteristics, treating effects, magnetic sedimentation efficiency and floc properties

Ming Zhang, a,b Feng Xiao a,⇑, Dongsheng Wang a,⇑, Xuzheng Xu a, Qi Zhou b

a State Key Laboratory of Environmental Aquatic Chemistry, Research Center for Eco-Environmental Sciences, CAS, Beijing 100085, PR China
b State Key Laboratory of Pollution Control and Resources Reuse, Key Laboratory of Yangtze River Water Environment, Institute of Biofilm Technology, College of Environmental Science and Engineering, Tongji University, 1239 Siping Road, Shanghai 200092, PR China

ARTICLE INFO

Article history:
Received 19 December 2016
Received in revised form 13 March 2017
Accepted 16 March 2017
Available online 18 March 2017

Keywords:
Ferromagnetic nanoparticles
Magnetic polyaluminum chloride
Magnetic seeding coagulation
Magnetic sedimentation
Floc property

ABS TRACT

The present work explored the ferromagnetic nanoparticle-seeded polyaluminum chlorides (PACls), and comparatively investigated their coagulation behaviors and magnetic sedimentation efficiency with the commonly adopted magnetic seeding coagulation. Three magnetic PACls were prepared by dosing ferromagnetic nanoparticles Fe3O4 or Fe(0) into the PACl (basicity 1.5) solution. Large clusters of Al species-magnetic nanoparticles existed as Alun in the magnetic PACl, exhibiting a colloidal surface configuration, according to the results of 27Al nuclear magnetic resonance and atomic force microscope. Compared with seeding magnetic nanoparticles into the treating influent, the magnetic PACls could lead to faster floc sedimentation and lower residual turbidity at equivalent doses of ferromagnetic nanoparticles and PACl; meanwhile, the DOC removal efficiency remained almost at the same level. The flocs produced by the superparamagnetic Fe3O4-involved PACl were less resistant for breakage and more difficult to re-form than those derived from the traditionally magnetic coagulation. Thus, after breaking the former flocs in the absence of external magnetic field, those separated magnetic seeds can be recovered via re-placing the broken flocs in the magnetic field. Importantly, the mass ratio of magnetic powder to coagulant in the magnetic coagulants was reduced from 10, 67, 100 or even higher to only 1.5–2.5. In sum, with respect to high magnetic separation efficiency, sound DOC removal, and low magnetic powder demand, the magnetic PACl with 50 mg/L Fe3O4 was considered as the preferable coagulant and the related coagulation-magnetic sedimentation process was more robust than the traditionally magnetic coagulation.

1. Introduction

To improve the treatment efficiency of coagulation-sedimentation, efforts have been dedicated to enhancing the floc formation, reducing the hydraulic retention time, increasing the overflow rate, and decreasing the system footprint [1,2]. In this context, the microcarrier-weighted coagulation is developed. Particularly, the ferromagnetic-weighted coagulation aided process, being named as magnetic coagulation, is growing in popularity at removing either non-magnetic or weakly magnetic matters from water [3,4]. The application of magnetic coagulation has been extended to the treatment of drinking water [5], surface water [6,7] and industrial wastewater especially in eliminating turbidity [8], heavy metal ions [9,10], dyes [11] and so forth.

The basic idea of magnetic coagulation-separation is “seeding” the strongly magnetic particles into a fluid suspension where they coagulate with target pollutants; the resulting “seed-pollutant” agglomerates are subsequently removed from the suspension either by sedimentation or filtration in the applied magnetic field [12]. One common way of magnetic seeding is to add magnetic components directly into the target water, and such practice is abundant. This process was used to remove color, turbidity, metals, organic matter and bacteria from waters [13–15]. Wan et al. compared the treating efficiency between two-stage dosing and one-stage dosing during the rapid mixing phase, in which magnetic seeds (FeO+Fe2O3 (Fe3O4)) and coagulant were added separately and simultaneously [16]. The former was found to perform better
than the latter. The leading mechanisms of the magnetic seeding process could be either the electrostatic attraction or the hydrophilic interaction [17,18]. Moreover, the magnetic seeding flocculation as well as the magnetic separation units were confirmed to take great advantages in eliminating turbidity and metal ions from surface water and drinking water [19,20].

Another novel and significant approach of magnetic seeding is to prepare magnetic coagulants by combining coagulants with ferromagnetic seeds. In the study of Liu et al. [21,22], the acidi treated chitosan and the fly ash were used to compound with FeO₄, respectively, forming magnetic coagulants to treat the algal blooms in a magnetic field. Zhang et al. prepared the magnetic polyanaluminum chloride (MPACL) by adding different ferromagnetic nanoparticles into the PACI solutions. The “Al species-nanoparticle” cluster was proposed to elaborate the unique properties of MPACL floc [23]. Additionally, given that the flocs resulted from the complex of ferric salts and polycarboxylic acid are more stable and easier to be removed at the magnetization conditions [24], the iron-based complex may also be considered as magnetic coagulants. Magnetic seeds-involved coagulants take great advantages in improving the water treatment efficiency and effect. Almost 99% of algal cells could be well removed by magnetic coagulants; meanwhile, residual chemical oxygen demand, total nitrogen, and phosphorus were controlled to a low level [21,22].

Seeding ferromagnetic particles in coagulants could obviously reduce the organic pollutants with low and mid-molecular weight as well as the biopolymers with high molecular weight compared with non-magnetic coagulation; therefore, the membrane fouling in the following filtration stage was significantly alleviated [25]. However, to the authors’ knowledge, there has been no specific study to compare the performance and potential of these two types of magnetic coagulation-sedimentation.

Humic acid (HA), an important hydrophobic natural organic matter (NOM), has always been regarded as a representative organic pollutant in scientific reports of water treatment [26,27]. HA has a high affinity to magnetic nanoparticles [28–31]: (i) the large specific surface area of nanoparticles offering space for HA adsorption; (ii) the Coulombic interaction between oppositely charged partners of HA and the magnetic surface; (iii) the ligand exchange between HA and the magnetite surface. Accordingly, it is preferable to use magnetic nanoparticles in the HA (or NOM) removal, and thus the magnetic separation could be facilitated.

This study aims at comparing the magnetic coagulant involved coagulation with the traditional magnetic seeding coagulation, and thus determining a promising way of magnetic coagulation. First, PACI and magnetic PACIs were comparatively characterized, and the effects of ferromagnetic nanoparticles on PACI properties were explored. Then, the comparison between MPACI-involved-coagulation (MC) and magnetic seeding coagulation (MSC) were conducted with respect to magnetic sedimentation efficiency, DOC removal, ë-potential variation, and floc properties. The two processes were elaborated from the standpoint of coagulation mechanisms.

2. Materials and methods

2.1. Chemicals

FeCl₃·6H₂O, FeCl₂·4H₂O, NaHCO₃, and absolute ethanol were obtained from National Medicines Corporation Ltd. of China. NH₃·H₂O, NaOH and HCl were provided by Beijing Chemical Plant. AlCl₃·6H₂O, NaNO₃ and Kaolin were provided by Xilong Chemical Corporation Ltd. of Shantou, Jinke Fine Chemical Institute of Tianjin, and Dongxu Chemical Plant of Beijing, China, respectively. Humic acid was produced by Sigma-Aldrich Corporation Ltd. of USA. All used reagents were analytical grade chemicals. Deionized water with conductivity lower than 0.5 mS/cm was used to prepare all the solutions and water samples.

2.2. Synthesis and characterization of magnetic nanoparticles

Two kinds of magnetic nanoparticles were used in this study: the laboratory-prepared Fe₃O₄ (LPF) and the superfine iron powder Fe(0) (SIP). The synthesis of LPF was based on the study of Hong et al. [32], and the specific procedure has been depicted previously [23]. The LPF product was dried at –50 °C under vacuum for 12 h. SIP was offered by the Research Center for Nano Technology of the Chinese Iron and Steel Institute.

The crystalline structures and sizes of LPF and SIP were examined by powder X-ray diffraction (XRD, D/Max-RC, Rigaku, Japan) using Cu Kα radiation. Vibration sample magnetometer (VSM; Lake Shore 7410), undertaken by the National Institute of Metrology of China, was used for the magnetic hysteresis curves of LPF and SIP in the magnetic field up to 20 kOe. The surface charge of magnetic nanoparticles was analyzed with a ë-potential meter (Zetasizer, Malvern, UK), during which the pH of suspensions was adjusted from 3 to 11 by adding NaOH or HCl solutions.

2.3. Preparation and characterization of PACI and MPACI

2.3.1. Preparation

The synthesis of PACI was similar to that described by Xu et al. [33], and the molar ratio of OH/Al (basicity value) was controlled to 1.5. PACI1.5 was then mixed with LPF or SIP in a shaking table at room temperature (25 °C) for 2 h at the speed of 250 rpm to prepare MPACIs. (PACI + LPF₂₅ mg/L)1.5 and (PACI + LPF₅₀ mg/L)1.5 were prepared by dosing 25 mg/L and 50 mg/L LPF into the PACI solution, individually, while (PACI + SIP₁₀₀ mg/L)1.5 was achieved by dosing SIP into PACI at the concentration of 100 mg/L. Those loadings were determined in view of the apparent dispersibility. The final composite MPACI1.5 s were aged 24 h, and conserved at 4 °C. For MC, all the MPACI1.5 s were ultrasonically vibrated for 15 min before dosing in order to well disperse the composite system.

2.3.2. Characterization

2.3.2.1. Liquid²⁷Al nuclear magnetic resonance (NMR). All coagulants were aged for one week, and then Al species of MPACI1.5s and PACI1.5 were measured by 500 MHz²⁷Al NMR. Instrumental settings and experimental conditions were addressed in the study of Xu et al. [33]. Mononuclear Al (Alm) and Al₁₃ were represented by signals near 0 and 62.5 ppm, respectively, and their fractions were calculated from the integral intensities of corresponding resonance peaks. Alun, the amount of undetectable species, was then obtained by:

\[ A_{un} = A_{l} - A_{m} - A_{13} \] (1)

2.3.2.2. Atomic force microscopy (AFM). AFM imaging was performed for the structural conformations of PACI1.5 and MPACI1.5s. The used AFM was a NanoScope IIIa Multimode Scanning Probe Microscopy Instrument (Digital Instruments, Santa Barbara, CA) equipped with an E-scanner operating in air at room temperature. The tapping mode AFM was employed to avoid sample destruction. The silicon cantilevers with 125 µm in length and 300 kHz in frequency (Nanoprobe, Model RTESPI4) were used. The cleaning process of mica plates was the same as that used by Ge et al. [34]. Two mica sheets were prepared and at least three images were obtained for each mica sheet sample. Images were taken with online
filtering and subsequently by flattening to remove the background slope. Using NanoScope Image analysis software, section analysis of images was conducted for the size estimation.

2.3.2.3. Scanning election microscopy and X-ray energy dispersion (SEM-EDX). The surface morphology and element composition of PAC11.5 before and after being compounded with ferromagnetic nanoparticles was characterized by SEM (LEO1530, Germany) coupled with an EDX detector. The sample was dried by vacuum freeze-drying for approximately 48 h at the vacuum degree of 55 Pa and the cold trap temperature of −50 °C, and then sputter coated with gold (Au) before being analyzed at 15 kV accelerating voltage. Principal components were extracted from the X-ray maps using Noran 7 COMPASS software.

2.4. Water sample

A synthetic water was prepared at room temperature (16 °C) by dissolving the HA stock solution and kaolin stock suspension in the de-ionized water with NaHCO₃ to provide ionic strength and pH buffer. In the synthetic water, average values of ζ-potential, turbidity, DOC, ionic strength and alkalinity were measured to be −54.4 mV, 79.5 NTU, 0.264 cm⁻¹, 2.239 mg/L, 1.0 mmol/L and 80 mg/L, respectively.

2.5. Coagulation and magnetic separation

Firstly, in the absence of external magnetic field, a series of jar tests were performed in a dosage range of 0.02–0.20 mmol/L as Al and an initial water pH range of 4–11. The related procedure and apparatus of jar tests can be referred to the former publication [23]. Thus, the PACI and MPACI dosages and initial pH condition were predetermined for the subsequent coagulation-magnetic separation trials. MC and MSC were then compared: after jar tests, all coagulated suspensions were immediately transferred to an external magnetic field of 2 T for sedimentation, and the turbidity of supernatant was measured at 5 min interval during the magnetically settling period by turbidimeter (Hach 2100P Turbidimeter, USA). The strong magnetic field was generated by a multi-pole permanent magnet (DLSM-II-4.0, Dawning Environmental Science and Technology Co., Ltd., China). It should be noted that, for MSC, aqueous suspensions of ferromagnetic nanoparticles were prepared by dispersing approximately 10 mg LPF or SIP nanoparticles into 100 mL de-ionized water and ultrasonically vibrating the mixture for 2 h. Those suspensions were conserved at 4 °C and used up in 2 days. In the jar test process, they were ultrasonically vibrated again for 15 min for the uniform dispersion before dosing. The MSC was carried out by simultaneously dosing PAC11.5 and the dispersion of ferromagnetic nanoparticles. Equivalent dosages of PAC11.5 and ferromagnetic nanoparticles were applied as in the MC tests.

A small amount of sample was taken immediately to measure the ζ-potential (Malvern, Zetasizer 2000, UK) after 30 s rapid mix in the coagulation stage. The sample of the supernatant water was taken for water quality analyses after the final sedimentation step (30 min). DOC of the supernatant was analyzed by a TOC analyzer (TOC-Vcph, Shimadzu, Japan) after filtration through 0.45 μm membrane, and pH values were measured using a pH meter (MP220, Mettler-Toledo, Switzerland).

2.6. Floc property

2.6.1. Floc formation, breakage and regrowth

A particle size analyzer (Mastersizer 2000, Malvern Corp., UK) with a small angle static light scattering was used to directly monitor dynamics of the floc growth-breakage-regrowth process. Details of the procedure can be found in the earlier publication [23]. Coagulation tests were carried out at the optimum coagulant dosage. Flocs formed by MC and MSC (viewed as “MC-flocs” and “MSC-flocs”, respectively) were compared from aspects of particle size, growth rate, strength factor and recovery factor. The strength factor (Sf) and recovery factor (Rf) can be calculated as follows:

\[ S_f = \frac{d_2}{d_1} \times 100\% \]  
\[ R_f = \frac{d_1 - d_2}{d_1} \times 100\% \]

where \(d_1\) is the average floc size of the steady region, \(d_2\) is the floc size after breakage, and \(d_3\) is the average floc size of the re-steady phase.

2.6.2. Fractal dimension (Df)

The fractal structure of flocs was investigated, and a higher Df value corresponds to a more compact interior floc structure [35]. Light scattering experiment is an important tool for its determination. The detailed principle and method has been presented by Jarvis et al. [36]. The light scattering technique involves measurement of light intensity \(I\) as a function of the scattering wave vector \(Q\), where \(Q\) is the difference between the incident and scattered wave vectors of the radiation beam in the medium. For independently scattering aggregates, the log–log plot of \(I\) against \(Q\) from data collected in scattering tests is linear, with a slope of \(D_f\):

\[ I \propto Q^{-D_f} \]

where the parameter \(D_f\) is the mass fractal dimension of the aggregates, ranging from 1 to 3 for physically relevant objects. A higher \(D_f\) value corresponds to a more compact interior floc structure [35].

3. Results and discussion

3.1. Properties of ferromagnetic nanoparticles

The diffraction characteristic peaks of LPF indicated its highly crystalline inverse cubic spinel phase; those of SIP corresponded to the diffraction planes of α-Fe (Fig. S1). Crystallite sizes of the LPF and SIP were calculated to be 11.2 nm and 9.3 nm according to the Scherrer’s equation [37]. It is generally accepted that the Fe₂O₃ and α-Fe particles turn to be superparamagnetic when their particle sizes are smaller than 16 nm and 5 nm, respectively [38]. Therefore, the LPF is qualified to be the superparamagnetic nanoparticle whereas the SIP should be of paramagnetism. The weak remanence effect of LPF (Fig. S2) confirmed that the LPF could show the superparamagnetic behaviors in an external magnetic field. For superparamagnetic materials, their thermal energy is able to overcome the anisotropy energy barrier of a single particle, and the net magnetization of particle assemblies in the absence of an external field is zero [39]. This property meets well with the requirement of magnetically assisted chemical separation [40]. Once the external magnetic field is removed after the magnetic sedimentation, no magnetic attraction exists between particles anymore; thus, the superparamagnetic particles may be more easily separated from the collected sludge through breaking the flocs; thereafter, magnetic particles can be recovered by replacing them in the magnetic field. Additionally, as shown in Fig. S3 the isoelectric point (IEP) of LPF was measured to be at approximately pH 6.4 while that of SIP was at around pH 6.6.
3.2. Characteristics of magnetic coagulants

3.2.1. Al species distribution and surface charge characterization

An increasing loading of magnetic nanoparticles resulted in the change of Al species distribution in coagulant (Table 1). The concentration of undetectable Al species rose whereas that of Al\(_{13}\), Al-monomer and Al-oligomer declined. Especially for PACl\(_{1.5}\) composited with the SIP at 100 mg/L, the Al\(_{un}\) content reduced to 37.7% while the Al\(_{13}\) and Al\(_{m}\) went up to 35.8% and 26.5%, respectively.

It is noteworthy that even a trace amount of magnetic nanoparticles (no more than 100 mg/L) could facilitate the formation of larger polymer such as the solid-phase Al\(_{un}\) or Al(OH)\(_3\). The large specific surface area, the inadequate coordination of surface atoms, and the strong adsorptive capacity of nano-scaled LPF and SIP might be the reasonable explanation for the aggregation in the MPACl\(_{1.5}\) solid-liquid system. As discussed previously [23], Al\(_{un}\) and Al\(_{13}\) possibly went through [Al\(_{m}\)] \rightarrow [Al\(_{un}\)] \rightarrow [Al\(_{un}\)] and [Al\(_{13}\)] \rightarrow [Al\(_{13}\)] \rightarrow [Al\(_{un}\)] in the compositing and aging process of MPACl. Here, [Al\(_{un}\)] was the aggregate of Al species and nanoparticles (presented as "Al species-nanoparticle cluster" in our study). The aggregates were of great potential in adsorbing-bridging, sweeping and enmeshing flocculation capacity. The \(\zeta\)-potential of PACl\(_{1.5}\) were close to zero, indicating the Al species (Al\(_{m}\), Al\(_{13}\) and Al\(_{un}\)) existing in the soluble form, while that of MPACl\(_{1.5}\) became positive, implying the adhesion of Al species on the ferromagnetic surface.

3.2.2. Morphological observation, size evaluation and element analysis

AFM was used to observe the morphological changes of PACl\(_{1.5}\) composited with ferromagnetic nanoparticles LPF and SIP. A spot-like photograph of PACl\(_{1.5}\) was found in Fig. 1(a), and there were two populations in width: smaller than 0.1 \(\mu\)m and 0.3–0.5 \(\mu\)m.

### Table 1
Species distribution and \(\zeta\)-potential of MPACl\(_{1.5}\) and PACl\(_{1.5}\).

<table>
<thead>
<tr>
<th></th>
<th>PACl(_{1.5})</th>
<th>(PACl + LPF(<em>{25\ mg/L}))(</em>{1.5})</th>
<th>(PACl + LPF(<em>{50\ mg/L}))(</em>{1.5})</th>
<th>(PACl + SIP(<em>{100\ mg/L}))(</em>{1.5})</th>
</tr>
</thead>
<tbody>
<tr>
<td>Al(_{un}) (%)</td>
<td>30.3</td>
<td>30.4</td>
<td>32.8</td>
<td>37.7</td>
</tr>
<tr>
<td>Al(_{13}) (%)</td>
<td>40.6</td>
<td>39.0</td>
<td>39.0</td>
<td>35.8</td>
</tr>
<tr>
<td>Al(_{m}) (%)</td>
<td>29.1</td>
<td>30.6</td>
<td>28.2</td>
<td>26.5</td>
</tr>
<tr>
<td>(\zeta)-potential (mV)</td>
<td>+2.5</td>
<td>+22.5</td>
<td>+28.6</td>
<td>+17.4</td>
</tr>
</tbody>
</table>

![Fig. 1. AFM images of coagulants: (a) PACl\(_{1.5}\), (b) (PACl + LPF\(_{25\ mg/L}\))\(_{1.5}\), (c) (PACl + LPF\(_{50\ mg/L}\))\(_{1.5}\), (d) (PACl + SIP\(_{100\ mg/L}\))\(_{1.5}\).](image)
On adding ferromagnetic nanoparticles, morphologies of MPACl1.5 s by AFM went different: the MPACl1.5 with 25 mg/L LPF was found to be sphere-like with 0.3–0.6 μm in width (Fig. 1(b)); at higher LPF concentration of 50 mg/L, larger aggregates with 0.8–2 μm in width appeared (Fig. 1(c)); the (PACl + SIP100 mg/L)1.5 exhibited multiple assembly structures with the largest size among investigated coagulants, including aggregates of sphere, branch-like structure and perforated sheets (Fig. 1(d)). Conformations and sizes of PACl1.5 and MPACl1.5 derived from AFM further confirmed the 27Al NMR analysis of Al species. It can be inferred that Al species congregated on ferromagnetic nanoparticles, forming “Al species-nanoparticle cluster” (Alun). The more amount of magnetic nanoparticles were added, the larger Alun could be achieved, leading to the conformation from small spots, large spheres to even larger aggregates.

The morphology of PACl before and after being mixed with ferromagnetic nanoparticles is related to the surface composition. Herein, SEM micrographs and EDX analysis of PACl1.5 and (PACl + LPF25 mg/L)1.5 are presented in Fig. 2 as representative of coagulants in this study. The inner structure of PACl1.5 was porous and irregular, containing several typical Al13 crystals (Fig. 2(a)); by adding LPF particles (Fig. 2(c)), the surface of MPACl1.5 turned to be compact with colloidal aggregates (Fig. 2(b)). The marked change could be attributed to the aggregation of Alun and Al13 on the LPF surface. Those Alun and Al13 aggregates assembled particles by interparticle bridging, led to the formation of Alun, and meanwhile gave rise to the colloidal configuration of MPACl. The element analysis and distribution images in Fig. 2(b) clearly demonstrated that blue spots (Fe) spread over the surface of MPACl sample we observed, revealing that magnetic particles had been well compounded with the MPACl coagulant.

3.3. Determination of coagulation conditions in the absence of external magnetic field

Preliminary coagulation tests were carried out to optimize the coagulant dosage and coagulation pH condition (Figs. 3 and 4). Initially, the pH value of the synthetic water was fixed at 7.60 ± 0.02 for the optimization of coagulant dosage. Residual DOC after coagulation decreased as the increase of coagulant dosage (Fig. 3(a)), and meanwhile the ζ-potential kept rising from negative to positive. The MPACl1.5 had a better organic matter removal at the dosage lower than 0.04 mmol/L as Al. The optimum dosage for organic matter removal (DOC removal) is 0.08 mmol/L as Al for both MPACl1.5 s and PACl1.5. Thereafter, results of pH optimization tests showed that the coagulation performance began to deteriorate when the initial pH was higher than 7.5. At pH of synthetic water higher than 10, MPACl1.5 s exhibited a stronger DOC removal capacity than PACl as shown in Fig. 4(a). The optimum coagulation condition for the DOC removal was determined as 0.08 mmol/L as Al for coagulant dosage and pH 7.00 ± 0.02 for the synthetic water, under which magnetic separation behaviors of MC and MSC from the aspect of residual turbidity in an external magnetic field were compared and discussed in following sections.

3.4. Coagulation-magnetic separation performance in the presence of external magnetic field

The residual turbidity of MC and MSC along with the settling time was comparatively shown in Fig. 5. It is evident that the final turbidity of both MC and MSC was lower than that of the coagulation without magnetic particles. MC yielded better turbidity removal efficiency than MSC for an equivalent ferromagnetic
nanoparticle dosage in a given magnetic field. For the LPF-involved magnetic coagulation-separation process, the higher LPF addition (50 mg/L) could give rise to lower residual turbidity for both MC and MSC. As for SIP, the difference between MC and MSC was smaller even though the highest addition of 100 mg/L SIP was used; the residual turbidity remained around 10 NTU, higher than that of LPF-involved processes. Hence, the preferable magnetic coagulation condition should be MC of (PACl + LPF50 mg/L)1.5 with respect to higher turbidity removal efficiency.

In the MSC, when being added into the synthetic water, those ferromagnetic nanoparticles would be negatively surface-charged (approximately −5.5 mV for LPF and −3.1 mV for SIP, according to Fig. S3) since the aquatic pH value was around 7.00. Thus, the highly positively charged PACl was more likely to interact with the negatively surface-charge kaolin particles by electrostatic attraction and binding affinity, forming flocs. As the ζ-potential of coagulated particles approached to zero and further reversed to be positive, the magnetic nanoparticles were apt to adsorb on coagulated flocs via the large specific surface area and high density of reactive surface. Additionally, the PACl that surrounded kaolin particles would separate magnetic seeds and kaolin particles, and reduce the opportunity of collision between kaolin particles and magnetic seeds [16].

In contrast, in the MC process, MPACls had been well combined with magnetic nanoparticles and Al species-magnetic nanoparticle clusters (considered as Al(III)) were preformed. On dosing into the synthetic water and mixed with kaolin particles, more magnetic nanoparticles could participate in the formation of flocs. Therefore, compared with MSC-flocs, the magnetic flocs formed by MC had greater potential of magnetic separation in an external magnetic field. Moreover, according to Fig. S3, the surface charge of LPF and SIP in MPACl should be positive since the pH of PACl1.5 was in the acidic range (lower than 4). Those positively charged nanoparticles could preferably agglomerate with the negatively charge contaminants, leading to the higher sedimentation efficiency of MC [4].

The comparison of MC and MSC in terms of DOC removal, ζ-potential and pH is presented in Fig. 6. The organic matter removal has been reduced from 2.239 mg/L to a quite low level (around 0.18 mg/L) by the non-magnetic coagulation-separation process. The residual DOC and pH value after MC and MSC did not make great difference under the optimum coagulating condition. It demonstrated that the trace amount of ferromagnetic nanoparticles in this study endowed the coagulated flocs a stronger sedimentation capacity by magnetic separation on the premise of keeping its capacity of eliminating the organic matter. The ζ-potential of coagulating systems by MC and MSC was slightly higher than that of the non-magnetic coagulation, confirming that the addition of ferromagnetic nanoparticles enhanced the attractive force between positively charged PACl and negatively surface
charged kaolin particles. On the account of the coagulation performance and the magnetic separation efficiency, MC with \((\text{PACl} + \text{LPF})_{1.5}\) is preferred in the magnetic coagulation-separation treatment.

### 3.5. Comparison between MC-flocs and MSC-flocs

When coagulating the synthetic water of HA and kaolin particles at a fixed coagulant dosage of 0.08 mmol/L as Al in MC and MSC processes, different floc growth-breakage-regrowth profiles were obtained, as presented by the median equivalent volumetric diameter \(d_{50}\) in Fig. 7. Floc properties including size, strength factor, recovery factor, and fractal dimension are summarized in Table 2.

In the growth period, both MC-flocs and MSC-flocs quickly reached their greatest size at the given shear rate. Coagulating suspensions stayed a rather short balance and then experienced a reduction of floc size, except the coagulation with \((\text{PACl} + \text{LPF}_{25 \text{mg/L}})_{1.5}\) and \((\text{PACl} + \text{SIP}_{100 \text{mg/L}})_{1.5}\) which kept the comparatively steady state in size. A general tendency illustrated that MC gave rise to smaller flocs. Subsequently, the floc breakage within the 1 min caused sharp drops of floc size to the same level (44–69 \(\mu\)m) in all coagulating suspensions. When the stirring speed turned back to 40 rpm, a reversibility phenomenon with regard to the floc size was observed, and MSC-flocs regrew to larger sizes in shorter time compared with MC-flocs. The regrowth plateau was always lower than that in the growth period indicating that the breakage of alum flocs under the stated conditions is not completely reversible [41].

It is apparent in Table 2 that LPF-involved MC promoted more compact flocs with lower strength factor than LPF-involved MSC. It has been well known that the floc strength is closely related to the particle bond strength and the number of interparticle bonds [42]. As interpreted in Section 3.2, compared with PACl, the increase of Al\(_{\text{un}}\) and the decrease of Al\(_{13}\) and Al\(_{\text{m}}\) in MPACls would strengthen their ability of bridging, sweeping and enmeshing pollutants, but somehow weaken the electrostatic attraction between MPACl and negatively charge surfaces. Thereby, the \((\text{PACl} + \text{LPF})_{1.5}\)-induced flocs were less shear-resistant than the \((\text{PACl}_{1.5} + \text{LPF})_{1.5}\)-induced. Compared with MSC with 100 mg/L SIP, stronger flocs were formed in the \((\text{PACl} + \text{SIP}_{100 \text{mg/L}})_{1.5}\)-involved MC. It may be attributed to the fact that the over-added magnetic nanoparticles (i.e. 100 mg/L in this study) was likely to aggregate themselves via the magnetic attraction, forming the shear-resist flocs.

For the MC-flocs, when exposing at high share rates, the Al species-magnetic nanoparticle clusters may be broken, and the

Fig. 5. Residual turbidity as a function of settling time at the predetermined coagulation condition.

Fig. 6. Coagulation performance of MC and MSC processes in the external magnetic field at the optimum coagulation condition: (a) Residual DOC, (b) \(\xi\)-potential and pH.

Fig. 7. Growth-breakage-regrowth files of flocs formed by MC and MSC.
rearrangement of residual Al species-magnetic nanoparticle clusters after breakage might occur, possibly hindering Al species interacting with the kaolin surface and reducing its ability to form bridges between particles. Therefore, all the MC-flocs generally showed the poorer recovery ability than the MSC-flocs. As denoted in Section 3.1, the LPF particles used in this work were superparamagnetic. If the external magnetic field is removed after the magnetic sedimentation, no magnetic attraction between particles will exist anymore. The weak LPF-involved MC-flocs may be easily destroyed by physical agitation whereas they are hard to re-

![Fig. 8. Possible mechanism of different magnetic coagulation processes: (a) formation of Al species-nanoparticle clusters in MPACl1.5, (b) coagulation-magnetic separation processes of MC and MSC.](image-url)

### Table 2

<table>
<thead>
<tr>
<th></th>
<th>$d_1$ (μm)</th>
<th>$d_2$ (μm)</th>
<th>$d_3$ (μm)</th>
<th>Strength factor $S_f$ (%)</th>
<th>Recovery factor $R_f$ (%)</th>
<th>Fractal dimension $D_f$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Growth</td>
<td>Breakage</td>
<td>Regrowth</td>
<td>Growth</td>
<td>Breakage</td>
<td>Regrowth</td>
<td>Growth</td>
</tr>
<tr>
<td>(PACl + LPF$_{25}$ mg/L)1.5</td>
<td>225.9</td>
<td>46.4</td>
<td>89.2</td>
<td>20.5</td>
<td>23.8</td>
<td>2.67</td>
</tr>
<tr>
<td>PACl1.5 + 25 mg/L LPF</td>
<td>239.6</td>
<td>51.5</td>
<td>145.8</td>
<td>21.5</td>
<td>50.1</td>
<td>2.64</td>
</tr>
<tr>
<td>(PACl + LPF$_{50}$ mg/L)1.5</td>
<td>212.5</td>
<td>48.4</td>
<td>124.4</td>
<td>22.8</td>
<td>46.3</td>
<td>2.65</td>
</tr>
<tr>
<td>PACl1.5 + 50 mg/L LPF</td>
<td>233.2</td>
<td>68.4</td>
<td>144.6</td>
<td>29.3</td>
<td>46.2</td>
<td>2.58</td>
</tr>
<tr>
<td>(PACl + SIP$_{100}$ mg/L)1.5</td>
<td>133.9</td>
<td>44.6</td>
<td>64.2</td>
<td>33.3</td>
<td>21.9</td>
<td>2.61</td>
</tr>
<tr>
<td>PACl1.5 + 100 mg/L SIP</td>
<td>248.6</td>
<td>52.4</td>
<td>130.5</td>
<td>21.1</td>
<td>39.8</td>
<td>2.66</td>
</tr>
</tbody>
</table>
form. Hence, the separated magnetic seeds can be collected and recovered via re-placing the broken flocs in the magnetic field.

3.6. Proposed mechanisms of MC and MSC

Based on the above discussion, MC and MSC performed differently in the coagulation-magnetic separation process. Feasible mechanisms of MPACl1.5 formation and coagulation-sedimentation of MC and MSC are schematically established herein. As illustrated in Fig. 8(a), Al species-nanoparticle clusters were formed when magnetic nanoparticles as added into PACl1.5 solution and the proportion of Al$_{\text{in}}$ was correspondingly increased. In that case, the magnetic component would sufficiently mix with coagulant and well participate in coagulation, forming flocs that were smaller and more compact due to the magnetic attraction between magnetic nanoparticles inside flocs. The flocs could easily precipitate in the presence of external magnetic field. However, when seeding magnetic nanoparticles directly into the target water (Fig. 8(b)), the traditional PACl-kaolin-HA flocs were apt to form. A part of magnetic nanoparticles would be wrapped into those flocs by particle collision and PACl coagulation. However, there would also be a small part of free magnetic particles which were not involved in the floc formation and finally separated by magnetic separation (called “ineffective magnetic seeds”). In that case, MSC-flocs would have lower sedimentation efficiency than MC-flocs.

Last but not least, the preparation and application of magnetic coagulants is promising with respect to greatly reducing the magnetic component dosage and thereby the cost of magnetic coagulation technique. On seeding magnetic components into wastewaters, the mass ratio of magnetic powder to coagulant was from 10, 67 to 100, or even higher [16,43]; for the application of magnetic coagulants, that ratio could be reduced to approximately 1.5−2.5 [21–23].

4. Conclusions

In the present work, the ferromagnetic nanoparticles seeded PACls were prepared as magnetic coagulants; their coagulation-magnetic sedimentation behaviors were then particularly compared with the magnetic seeding coagulation in an external magnetic field of 2 T. The superparamagnetic LPF and paramagnetic SIP significantly influenced the Al species, the morphology as well as the size of Al species. By adding LPF and SIP, the content of large-size [Al$_{\text{in}}$] in MPACl increased. The formation of “Al species-nanoparticle clusters” led to the colloidal configuration of MPACl.

In the external magnetic field, the MPACI-involved magnetic separation could result in a lower residual turbidity and a higher magnetic sedimentation efficiency compared with the magnetic seeding coagulation. The MPACI exhibited a high organic matter removal efficiency compared with PACI alone. MC-flocs were smaller, weaker and less reversible than MSC-flocs. Particularly, the weak LPF-involved MC-flocs may be easily destroyed by physical agitation whereas they are hard to re-form. Those separated magnetic seeds are highly possible to be recovered via re-placing the broken flocs in the magnetic field. Considering the coagulation-separation performance and floc properties, (PACl + LPF50 mg/L)$_{1.5}$ should be favored by the practical application. The MC-separation was superior to the MSC-separation process because of the much lower demand of magnetic particles and the better settleability.

Acknowledgements

This research was funded by the following foundations: National Natural Science Foundation of China (No. 51608373, 51378014, 51338008, 51338010, 21107125), 59th China Postdoctoral Science Foundation (No. 2016MS591713), and Foundation of State Key Laboratory of Pollution Control and Resource Reuse (Tongji University), China, (No. CRRY15001). The authors are very grateful to the technical support from the Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences.

Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.seppur.2017.03.028.

References

[38] R.Y. Hong, Preparation and application of magnetic nanoparticles and fluid, China, 2008.