Investigation of antibiotics in mollusks from coastal waters in the Bohai Sea of China

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1. Introduction

Antibiotics are widely used to treat infectious diseases for both humans and animals (Kümmerer, 2009). Some antibiotics are also used in livestock farming, aquaculture and agriculture (Cruz Moreno-Bondi et al., 2009). However, many antibiotics cannot be completely metabolized in the body, and the excreted portion cannot be eliminated by the sewage treatment plants. In addition, direct disposal of medical and industrial wastes also contain antibiotics. Therefore, the water cycle is contaminated with a large amount of antibiotics (Massey et al., 2010). They persist in the environment, because most of them are replaced by ongoing wide use, although they are being degraded at a certain rate (Cunningham et al., 2009).

Antibiotic contamination has attracted increasing attention due to its undesirable effects on ecosystems and the health of humans and animals (Sapkota et al., 2008; Wang, 2009; Ye et al., 2007). Most studies have focused on the antibiotics that may cause resistance among natural bacterial populations. In recent years, more and more attention was paid to the antibiotic resistance genes (ARGs), which have been recognized as a new emerging contaminant in environment (Luo et al., 2010; Zou et al., 2011). It has been shown that not only this resistance can be transmitted to the general population that facilitates antibiotic-resistant infections, but also antibiotic-resistant bacteria can pass through the food chain from animals to humans (Canada-Canada et al., 2009; Díaz-Cruz et al., 2009; Sapkota et al., 2008).

As the most common antibiotics, quinolones (QNs), sulfonamides (SAs) and macrolides (MCs) have been found in different environmental compartments, such as water (Ternes et al., 2007), sewage sludge (Golet et al., 2002), sediments (Arikan et al., 2008), and soils (Hu et al., 2010; Tantam et al., 2011), and in foods (Nie et al., 2009) and animals (Juan-Garcia et al., 2007). An increasing number of analytical methods have been developed for animals and foods, while only very limited information is available for the occurrence and fate of antibiotics, especially for marine system such as gulf, bay or sea, where many pollutants have been concerned to a great extent due to their transfers to open sea (Zhang et al., 2009).

Bohai Bay is the biggest semi-enclosed sea located in the northeast of China and the Bohai Bay Economic Rim surrounding Bohai Sea is an important economic center that possesses about 17% of the total population and contributes about 25% of total GDP of China (Wang et al., 2008). Rapid economic development has been associated with serious environmental pollutions problems. Many pollutants have been proved to be widespread in Bohai Bay in the previous study (Liang et al., 2003; Wang et al., 2007). However, research on antibiotic pollution in Bohai Bay area has been very limited (Na et al., 2009; Zou et al., 2011).
Recently, several studies have used mollusks as potential bio-monitors because of their high accumulation capacity and high abundance in marine ecosystems (Pan et al., 2010; Yang et al., 2008). Therefore, we used mollusks in this study to investigate the distribution of quinolones, sulfonamides, and macrolides from nine typical coastal sites along the Chinese Bohai Sea. This study is the first to demonstrate the ubiquitous occurrence of three major types of antibiotics in a large water system.

2. Materials and methods

2.1. Regents

HPLC-grade methanol and acetonitrile were purchased from Fisher Scientific (Pittsburgh, PA, USA); Ammonium formate (99%) and ammonium hydroxide (V/V, 50%) were purchased from Alfa Aesar (USA); Formic acid (98%) was purchased from Fluka (USA). All purity water (>18.2 MΩ cm) was prepared with Milli-Q Advantage A10 system (Millipore, USA).

Ofl oxacin (OFL, 99.9%), norfloxacin (NOR, 99.9%), ciprofloxacin (CIP, 99.9%), sarafloxacin (SAR, 95.0%), flor oxacin (FLE, 99.5%), lomefoxacin (LOM, 98.0%), difloxacin (DIF, 98.0%), enrofloxacin (ENR, 99.9%), sulfadiazine (SZD, 99.7%), sulfamerazine (SMR, 99.9%), sulfamethoxine (SMX, 99.4%), sulfacetamide (STA, 99.0%), sulfamonomethoxine (SMM, 99.0%), erythromycin (ERY, 99.1%), roxithromycin (ROX, 90.0%), josamycin (JOS, 98.0%), tylosin (TYL, 82.4%), and spiramycin (SPI, 88.9%) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Sulfathiazole (ST, 99.0%), sul fyapyrimidine (SPD, 95.0%), sulfamethoxazole (SMX, 99.0%), and sulfamethazine (SDM, 99.0%) were purchased from Kasei Industry Co., Ltd. (Tokyo, Japan).

The following isotopically labeled compounds were used as surrogates standards (1000.0 μg/L in methanol): norfloxacin-d₃ (NOR-d₃), ofloxacin-d₃ (OFL-d₃) and sarafloxacin-d₄ (SAR-d₄) that were purchased from Sigma–Aldrich (St. Louis, MO, USA); and sulfamethoxazole-d₄ (SMX-d₄), sulfamethazine-d₄ (SDM-d₄), spiramycin-d₁ (SPI-d₁), and erythromycin-13C₃, d₁ (ERY-13C₃, d₁) that were purchased from Toronto Research Chemicals (Oakville, ON, Canada).

2.2. Sample collection

The samples were collected three times in late July to early August of 2006, 2007 and 2009. A total of 190 pooled mollusk samples were collected from nine coastal cities along the Bohai Sea, including Dalian, Yingkou, Huludao, Beidaihe, Tianjin, Shouguang, Penglai, Yantai and Weihai (Fig. 1). Most of the sampling sites were large and busy seaports with a large population. The population in the selected cities was shown in Table S1. Eleven species of mollusks that are commercial seafoods were selected, including Neverita didyma (Nev), Amussium (Amu), Meretrix meretrix (Mer), Mactra veneriformis (Mac V), Mactra chinesis (Mac C), Mya arenaria (Mya), Neverita didy ma (Nev), Rapana venosa (Rap), and Mytilus edulis (Myt) (Table S2).

All mollusk samples were always kept at low temperature after collection. They were unshelled in the laboratory, and tissue homogenate was lyophilized and ground into fine powders before analyzed. All samples were stored at −20 °C until chemical analysis.

2.3. Sample preparation

Chemical analysis was following the EPA method 1694 developed by the US Environmental Protection Agency (EPA) (EPA, 2007) with some modifications. The twenty-two target compounds in mollusk samples, including eight quinolones, nine sulfonamides, and five macrolides, were simultaneously extracted using an ASE 350 pressurized liquid extraction (PLE) system (Dionex, Sunnyvale, CA, USA). 0.1 g of pretreated lyophilized mollusk sample was spiked with 20 ng of internal standard (Nor floxacin-d₃, Ofloxacin-d₃, Sarafloxacin-d₄, Sulfamethoxazole-d₄, Sulfamethazine-d₄, Spiramycin-d₁, and Erythromycin-13C₃) before being thoroughly mixed with 7 g of diatomite from Varian (Walnut Creek, CA, USA). The mixture was put into a 34 mL stainless steel extraction cell with 100% methanol as the extracting solvent.

The extraction conditions were: extraction temperature, 70 °C; extraction pressure, 1500 psi; preheating period, 5 min; static extraction, 10 min; final extraction volume, 60 mL; flush volume 60% of the cell volume; nitrogen purge, 120 s; and number of extraction cycles, 2. Each PLE extract was concentrated by RE-2000 rotary evaporator (Yarong, Shanghai, China) to a final volume of about 1 mL at 37 °C and 0.08 MPa in 100 mL round-bottom flasks. Immediately after concentration, the extract was transferred to a 100 mL conical tube and the round-bottom flasks was rinsed twice with 0.5 mL of methanol, and then add 100 mL of regent water to this conical tube.

Solid-phase extraction was performed on 6 mL Oasis HLB sorbent cartridges (200 mg; Waters) that were pre-conditioned with 5 mL methanol and 5 mL water at a flow rate of 1 mL/min. After they passed through the cartridges, the samples were washed with 10 mL of water and vacuum-dried for 30 min. Analytes were eluted with 5 mL methanol containing 5% (v/v) ammonia into 15 mL conical tube flask. The eluate was collected and concentrated to 1 mL under a gentle stream of nitrogen before being dissolved with 1 mL of a mixture of methanol/aqueous solution of formic acid, 0.05% (10:90 v/v). Then the extract was centrifuged for 5 min at 12,000 rpm. Finally, the supernatant was filtered through a 0.22 μm nylon filter, and an aliquot (15 μL) of it was injected for liquid chromatography/tandem mass spectrometry.

2.4. Liquid chromatography/tandem mass spectrometry

The LC-MS/MS system consisted of an Ultimate 3000 HPLC (Dionex, Sunnyvale, CA, USA) and a triple-quadrupole mass spectrometer (API 3200; Applied Biosystems/MDL SCIEX, US) with electrospray ionization (ESI). Xterra MS C₁₈ column (3 μm, 100 mm × 2 mm) was used as the analytical column at a flow rate of 0.20 mL/min. Methanol-acetonitrile (1:1, V/V) were used as mobile phase A, and 0.3% formic acid/water (containing 0.1% ammonium formate, V/V, pH = 2.9) was used as mobile phase B. The gradient program was as follows: the mobile phase starting conditions were 10% of A for 2.0 min, and A was increased to 70% in 10.0 min before being increased to 100% in 4.0 min; 100% of A for 3.0 min, followed by returning to the initial composition in 0.1 min, which was maintained for 13.9 min. The total run time was 33.0 min.

For the MS detection, the instrument was operated in the positive electrospray ionization and multiple reactions monitoring (MRM) mode. The MS/MS parameters are as follows: curtain gas pressure, 0.14 MPa; collision gas pressure, 0.02 MPa; ion spray voltage, 5000 V; Temperature, 600 °C; Gas1, 0.38 MPa; and Gas2, 0.45 MPa. Other parameters of MS/MS and ion pair are listed in Table S3.

Fig. 1. Sampling locations of mollusk samples along the Bohai Sea in China.
2.5. Quality assurance and quality control

The concentrations were determined by an internal standard method. Ten concentration points (0.01, 0.05, 0.1, 0.5, 1, 5, 10, 50, 100, and 500 μg/L in water–methanol (9:1, v:v)) of individual antibiotics were used as standards, and the linearity of calibration curves was confirmed (r² > 0.99).

The degradation loss during storage was determined as not significant (<20%) by measuring the antibiotic concentrations before and after one month’s storage in the dark at −20 °C. Field blanks and procedural blanks were analyzed with extraction to control travel contamination and laboratory contamination, and all equipments and containers were rinsed with methanol and water before use to avoid analytical interference and/or cross contamination.

The limits of detection (LODs) and limits of quantification (LOQs) are listed in Table S4. LODs and LOQs were determined as the amounts for which signal-to-noise ratios (S/N) were higher than 3 and 10, respectively. A 10 μg/L standard was set as the quality control concentration, which was checked every 10 injections to ensure analysis stability and to verify calibration. The calibration curve was used for quantification only when the quality control standard was within 20% of its initial value.

2.6. Statistical analysis

All statistical analyses were performed with IBM PASW Statistics 18.0 (SPSS Inc., 1993–2007). One-way analysis of variance (ANOVA) and principal component analysis (PCA) were used to visualize the relationships among the antibiotic concentrations in the experimented mollusks. It was considered as statistically significant difference when p < 0.05 for ANOVA. For PCA, principal factors should account for approximately 75% of the total variance, and the principal components (PCs) were extracted for eigen values that were greater than 1 (Wiechula et al., 2006).

3. Results and discussion

3.1. Antibiotics in mollusks

The dry-weight-based concentrations of 22 antibiotics, including quinolones, sulfonamides and macrolides, are listed in Table S5. All target antibiotics except tylosin were detected in the 190 mollusks samples. QNs were the most concentrated, with a mean ∑QNs concentration of 86.76 μg/kg. The levels of SAs and MCs were lower than QNs', with mean concentrations of 5.99 and 2.58 μg/kg, respectively. The concentrations of individual QNs, SAs and MCs are illustrated by box-and-whisker plots in Fig. 2b, c and d, respectively. NOR, CIP, FLE, OFL and SAR were the predominant antibiotics, with higher than 10.00 μg/kg of mean concentrations and high detection frequencies (>75%). As a contrast, SDZ, SDM, SDMD, JOS and ROX had lower than 0.20 μg/kg of mean concentrations and low detection frequencies (<40%).

3.1.1. QNs

The prominent antibiotics in the mollusks were QNs. The concentration of ∑QNs ranged from 8.79 (Mer, Yantai) to 557.00 μg/kg (Sca, Yantai) in the 2006 samples, from 0.71 (Amu, Weihai) to 1575.10 μg/kg (Mer, Dalian) in the 2007 samples, and from 3.93 (Rap, Yingkou) to 398.00 μg/kg (Mac C, Weihai) in the 2009 samples, respectively. The detection rate of eight quinolones ranged from 56 to 87%, indicating extensive quinolones pollution along the Bohai coast (Table 1). Compared with SAs and MCs, quinolones were strongly adsorbed into sewage sludge and sediments without biodegradation, which promotes their persistence in the environment (Cruz Moreno-Bondi et al., 2009). In addition, quinolones were stable in edible animal tissues (Juan-Garcia et al., 2006).

The mean concentrations of QNs in the mollusks were in the order of “NOR > OFL > CIP > FLE > SAR > LOM > ENR > DIF”. The concentrations of NOR ranged from below detection limit (BDL) to 370 μg/kg (mean 18.82 μg/kg), followed by those of OFL (BDL to 242 μg/kg, mean: 14.65 μg/kg), CIP (BDL to 208 μg/kg, mean: 14.65 μg/kg), OFL (BDL to 333 μg/kg, mean: 11.16 μg/kg), and SAR (BDL to 333 μg/kg, mean: 11.16 μg/kg). The
SAs had been found previously in mollusks in this region. Na et al. (2009) reported high concentration of SDZ in Mac C of Dalian in China (6.5–143.3 µg/kg), but no detectable SMM and SMX. However, similar concentrations of SAs (SMR, 0.35–0.39 µg/kg, SD, 0.32–0.78 µg/kg, SMX, 0.95 µg/kg) as those in the present study in hatcheries and mussels (Myt) were detected in Spain (Fernandez-Torres et al., 2010). Compared to other foodstuffs, the concentrations of SAs in mollusks from the Bohai Sea were lower. Higher concentration of SDMD (1.1–43.2 µg/kg) and SDZ (2.0–160.9 µg/kg) in swine muscle tissues was reported in Spain (Chico et al., 2008), and higher level of SDZ (83 µg/kg) in bovine muscle tissues was also detected in Spain (Carretero et al., 2008).

### 3.1.3. MCs

MCs were only detected in a few mollusks with detection rates of between 1 and 65%. The concentration of \( \sum \)MCs ranged from BDL to 26.30 µg/kg (Nev, Beidaihe) in the 2007 samples, from BDL to 31.61 µg/kg (Mac C, Yingkou) in the 2007 samples, and from BDL to 28.55 µg/kg (Rap, Huludao) in the 2009 samples.

The concentrations of MCs were in the order of “ERY > SPI > ROX > JOS > TYL”. The major macrolides in mollusks were ERY, which had mean concentration of 1.65 µg/kg and accounted for 64.0% of \( \sum \)MCs. This is consistent with the fact that ERY is the most commonly used MCs in Bohai Bay. The concentration of SPI was the second highest with 1.65 µg/kg of mean level. ROX and JOS were only found in a few samples with 24 and 7% of detection rates, respectively. In addition, TYL was not detected in any sample.

Data on MCs in mollusks were also limited all over the world and information on other food was available. Similar concentration of ERY (3.2 µg/kg) and TYL (2.4 µg/kg) in bovine muscle tissues was reported in Spain (Juan et al., 2010), while higher concentration of ERY (8.6 µg/kg) and TYL (2.0–160.9 µg/kg) in swine muscle tissues were found in Spain (Vidal et al., 2009).

Up to now, limited data on the accumulation of antibiotics in mollusks is available. In recent study it was reported that the mean concentrations of individual QNs, SAs and MCs in Bohai Sea water were in the range of 110–460, 17–37 and 30–113 ng/L, respectively (Zou et al., 2011). We roughly analyzed the potential accumulation of antibiotics based on this data, indicating that most selected antibiotics may be slightly accumulated in the mollusks.

### 3.2. Annual variation of antibiotics

Box-and-whisker plots are shown in Fig. 2 to visualize the distribution of antibiotics in mollusks. No significant variation was found for the concentrations of \( \sum \)QNs, \( \sum \)SAs and \( \sum \)MCs among the three years (Fig. 2a). The results of analysis of variance (one-way ANOVA) for QNs, SAs and MCs are listed in Table 2. The levels of QNs, SAs and MCs were limited all over the world and information on other food was available. Similar concentration of ERY (3.2 µg/kg) and TYL (2.4 µg/kg) in bovine muscle tissues was reported in Spain (Juan et al., 2010), while higher concentration of ERY (8.6 µg/kg) and TYL (2.0–160.9 µg/kg) in swine muscle tissues were found in Spain (Vidal et al., 2009).

![Box-and-whisker plots](image)

**Table 1**

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Mean</th>
<th>Median</th>
<th>Minimum</th>
<th>Maximum</th>
<th>Detectable (%)</th>
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14.54 µg/kg), FLE (BDL to 250 µg/kg, mean: 13.57 µg/kg), and SAR (BDL to 160 µg/kg, mean: 10.94 µg/kg). The last three QNs, LOM, ENR, and DIF, were frequently detected but with low mean concentrations below 6.00 µg/kg.

QNs were rarely analyzed in mollusks around the world, while some data in other seafood species were available (Juan-Garcia et al., 2007; Zhao et al., 2007). NOR, OFL, CIP and ENR were often detected in fish samples. For example, NOR was found to be the main contaminant of Anguilla japonica tissues with a concentration of 2.0–100.54 µg/kg in fish from the Pearl River Delta, China (Nie et al., 2008), which is consistent with the present study. The high concentration of NOR is probably caused by its over-consumption in China, partially due to its low price. Xie et al. (2008) reported lower concentrations of OFL (1.5–7.4 µg/kg), CIP (7.6–31.6 µg/kg), and ENR (3.9–13.5 µg/kg) in roast eels in China. Compared with other foodstuffs, the concentrations of QNs in mollusks from the Bohai Sea were slightly high. Zhao et al. (2007) reported lower concentration of NOR (5.6–12.5 µg/kg), CIP (1.5–59.3 µg/kg) and ENR (17.3–85.3 µg/kg) in chicken muscle tissues in China, and similar concentration of NOR (1.1–43.2 µg/kg), CIP (1.0–73.6 µg/kg) and ENR (2.0–160.9 µg/kg) in swine muscle tissues.

#### 3.1.2. SAs

The concentrations of SAs in mollusks from Bohai Sea were lower than those of QNs. The levels of sulfonamides ranged from BDL to 69.50 µg/kg (Sca, Shouguang) in the 2006 samples, from BDL to 76.75 µg/kg (Nev, Beidaihe) in the 2007 samples, and from BDL to 14.41 µg/kg (Myt, Dalian) in the 2009 samples. Furthermore, the \( \sum \)SAs concentrations in mollusks in the Bohai Sea were all lower than the maximum residue limits of 100 µg/kg, according to European Commission, Council Regulation (EU) No 37/2010 (EU, 2009).

The concentrations of SAs were in the order of “SMM > ST > SMX > SIA > SMR > SPD > SDMD > SDZ > SDM”, and the first four sulfonamides in mollusks accounted for 81.2% of \( \sum \)SAs. SMM possessed the highest mean concentration of 1.6 µg/kg, followed by ST, SMX and SIA, with mean concentrations of 1.33, 1.01, and 0.91 µg/kg, respectively. The mean concentrations of SMR and SPD were only 0.42 and 0.26 µg/kg, respectively, although they were frequently detected in samples. The last three SAs were not frequently detected, and their mean concentrations were below 0.20 µg/kg.
caused by the prohibition of some SAs as feed additive for particular fish species, according to the Use Standard of Pollution-Free Food and Fish that was printed and issued by the Ministry of Agriculture in 2002.

3.3. Comparison of sampling sites

No significant differences were observed for SSSAs in mollusks among the nine sampling sites (Fig. S1a). However, higher residue levels of SQNs were detected in Dalian (mean 181.64 μg/kg), while these levels were much lower in Beidaihe (mean, 50.73 μg/kg) and Yingkou (mean, 56.48 μg/kg). Furthermore, higher concentrations of ΣMCs (mean, 4.11 μg/kg) were detected in Beidaihe, while they were low in Dalian (mean, 0.85 μg/kg). In general, the result showed that the pollution was antibiotics species dependent and widespread in this area with high population density.

PCA was performed to analyze the residual antibiotics with detectable rates of over 60%, which included seven QNs, two SAs, and one MC. The varimax rotation reduced all variables to three different PCs, which represent 75.58% of the total variance. According to these rules, loadings are situated closely in bitplots represent the variables that are highly correlated. Therefore, PCA was used to identify the correlation between various parameters of pollutants. As shown in Fig. 3, PC1 accounted for 53.96% of the total variance and had strong positive loadings on seven QNs; PC2 accounted for as much as 11.43% of the total variance and had moderately positive loadings for ERY and SPD; PC3 accounted for 10.20% of the total variance and had high correlation with SMM.

Dalian had higher loading values of PC1 in the component plot (Fig. 4), indicating that residual QNs were the main antibiotic pollutant of Dalian, which is one of the largest seaports in China. In addition, Beidaihe was highly polluted by ERY and SPD, and Huludao was severely contaminated by SMM (Fig. 4).

3.4. Comparison of mollusks species

No significant differences in ΣMCs concentrations were observed among different species of mollusks (Fig. S2a). However, relatively high residue levels of ΣQNs were detected in Mac V (mean 128.87 μg/kg) and Mer (mean 128.19 μg/kg), while low residue levels of ΣQNs were detected in Mya (mean 43.32 μg/kg) and Amu (mean 51.87 μg/kg). In addition, significantly higher levels...
of SAs were observed in Myt (mean 9.60 μg/kg) and Sca (mean 9.51 μg/kg) than those in Mac C (mean 1.77 μg/kg) and Rap (mean 2.93 μg/kg).

PCA was also applied to determine the characters of pollutants in different mollusks species. Mac V and Mer had higher loading values of PC1 and PC3 in the component plot (Fig. 5), indicating that the concentrations of QNs and SMM in these samples were relatively higher; and Myt had higher loading values of PC2, indicating much higher concentrations of ERY and SPD than other species.

3.5. Risk assessment for the seafood

Mollusks are one of the most popular seafoods in China, especially for those who live in coastal cities. Therefore, it is necessary to evaluate whether dietary intake of mollusks containing antibiotics is harmful to public health. Indeed, the EU has set MRLs for some residual antibiotics in foods of animal origin to protect public health, based on related scientific assessments (Table S6). In the present study, 15 out of 190 samples exceeded the MRL of SAR (30 μg/kg) for muscle of Salmonidae, and 5 of the 15 samples were from Dalian. As for ENR (Sum of enrofloxacin and ciprofloxacin), 8 out of 190 samples were above the MRL (100 μg/kg), 3 of which were from Dalian. Therefore, it is reasonable to advise that consumption of mollusks in the Bohai Sea may pose potential health risks to local residents. In addition, Dalian should have extra attention than other areas, especially for those who live in coastal cities. Therefore, it is necessary to evaluate whether dietary intake of mollusks containing antibiotics is harmful to public health. Indeed, the EU has set MRLs for some residual antibiotics in foods of animal origin to protect public health, based on related scientific assessments (Table S6). In the present study, 15 out of 190 samples exceeded the MRL of SAR (30 μg/kg) for muscle of Salmonidae, and 5 of the 15 samples were from Dalian. As for ENR (Sum of enrofloxacin and ciprofloxacin), 8 out of 190 samples were above the MRL (100 μg/kg), 3 of which were from Dalian. Therefore, it is reasonable to advise that consumption of mollusks in the Bohai Sea may pose potential health risks to local residents. In addition, Dalian should have extra attention than other areas, because it may have the highest antibiotic contaminations in mollusks.

4. Conclusion

In this study, the levels of residue antibiotics were investigated for various mollusks that were collected from nine coastal cities along the Chinese Bohai Sea. QNs were widely distributed in mollusks in this area. However, no significant changes were observed in the concentrations of QNs and MCs from 2006 to 2007 and to 2009, while those of SAs were reduced in this period of time. Furthermore, the levels of QNs were higher in Mac V and Mer than that in other species, indicating that these two species can be used to evaluate the contamination of QNs in the Bohai coastal areas.

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Appendix. Supplementary material

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.envpol.2011.10.022.

References


